# Neutralizing Interleukin-1 beta Protects Islet ß-cells From Intracellular Amyloid

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#### INTRODUCTION

- Type 2 Diabetes (T2D; adult-onset diabetes) is characterized by the progressive loss of pancreatic ßcell mass and function.
- Aggregation of the toxic protein, amyloid, contributes to the loss of ß-cell mass. Amyloid formation is also observed during pre-transplant islet culture and islet grafts in patients with type 1 diabetes (T1D).
- Amyloid formation contributes to islet inflammation by stimulating the production of the pro-inflammatory cytokine interleukin-1 beta (IL-1ß) in islets.

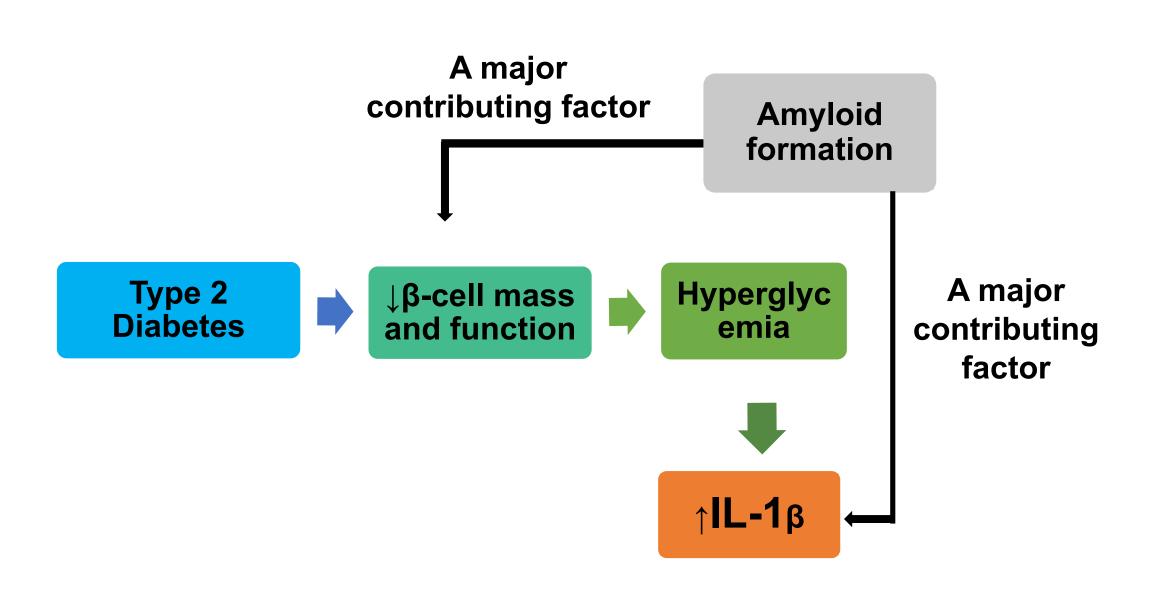


Fig 1. Pathogenesis of T2D.

• IL-1ß neutralizing monoclonal antibodies (nAb) are able to effectively block IL-1ß action by targeting IL-1ß.

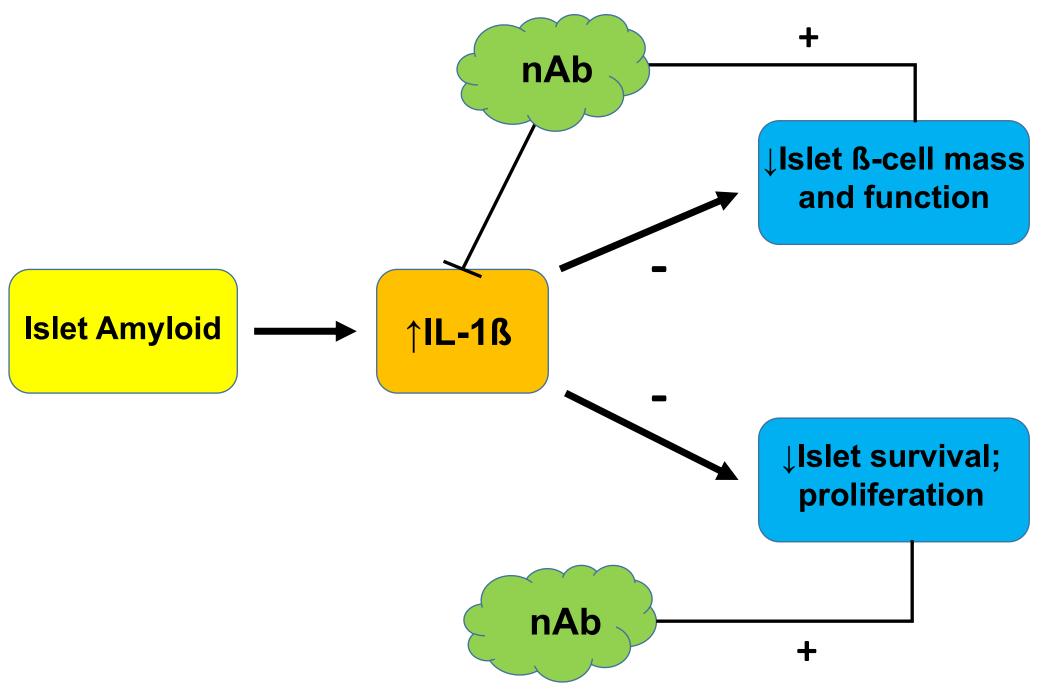


Fig 2. The mechanism of amyloid-induced ß-cell toxicity and proposed protecting mechanism of neutralizing antibody (nAb).

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#### AIMS

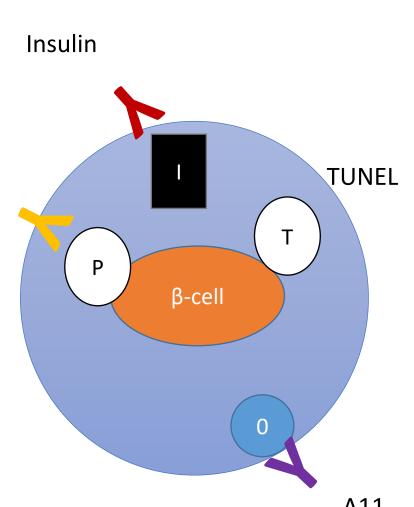
We examined if:

- Blocking IL-1ß signalling can reduce the intracellular amyloid-induced ß-cell death.
- 2. Blocking IL-1ß signalling can enhance ß-cell survival in the presence of intracellular amyloid.

#### **METHODS**

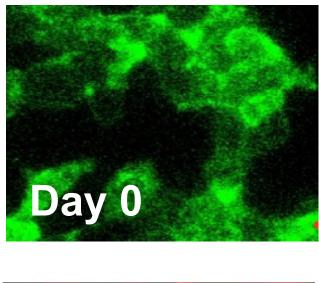
- INS-1 ß-cells (n=3 independent studies) were cultured in RPMI-1640 medium after transduction with prohIAPP-adenovirus to induce intracellular amyloid formation.
- INS-1 
  ß-cells were treated with nAb (1 µg/mL)
- Quantitative immunohistochemistry was performed on INS-1 ß-cells for insulin and A11 (small intracellular aggregates), TUNEL (apoptosis), or PCNA (proliferation).

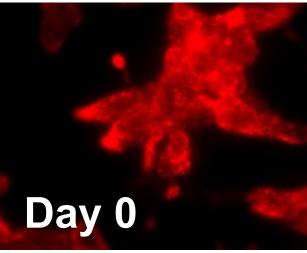
PCNA Fig 3. INS-1 ß-cells were immunolabelled for insulin and A11, TUNEL, or PCNA



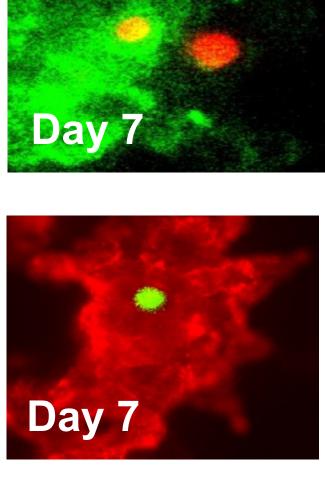
**Fig 5**. The proportion of TUNEL-positive (apoptotic)  $\beta$ -cells after 7-day treatment with nAb. Day 0 (C) and Day 7 (nontreated, NT) are shown for comparison. Data are expressed as mean±SEM of three independent studies, \*=p<0.05.

### RESULTS

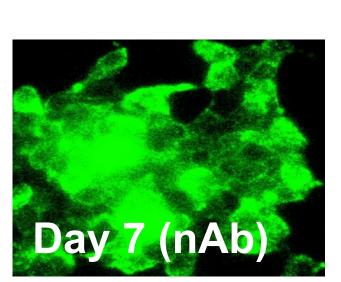


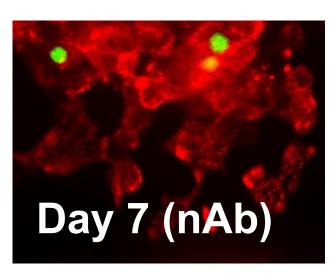


#### **INSULIN/TUNEL**



**INSULIN/PCNA** 

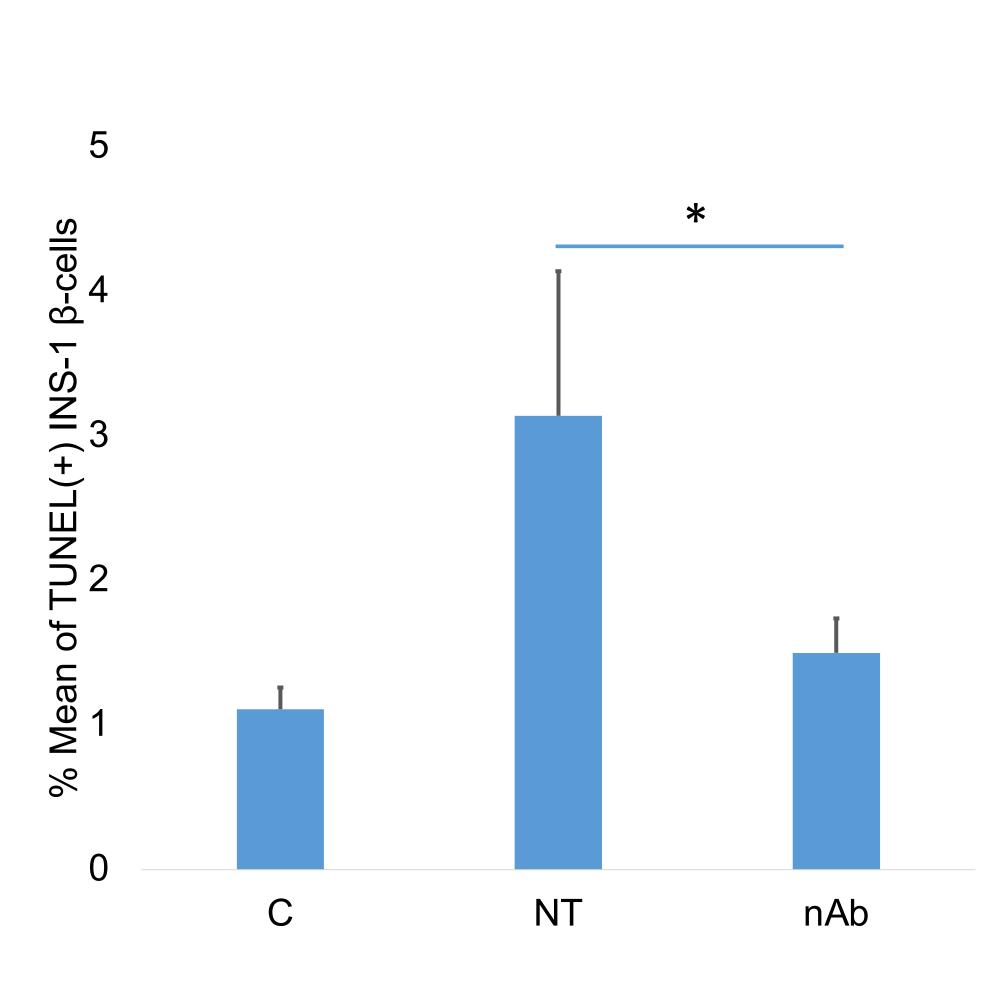




**Fig 6.** The proportion of PCNA-positive (proliferative) β-cells after 7-day treatment with nAb. Day 0 (C) and Day 7 (nontreated, NT) are shown for comparison. Data are expressed as mean±SEM of three independent studies, \*=p<0.05.

Fig 4. INS-1 ß-cells from control, non-treated and treated (with neutralizing IL-1ß antibody (nAb)) were immunolabelled for insulin and PCNA (top row) or TUNEL (bottom row). Micrographs represent three independent studies.

### **RESULTS**, continued



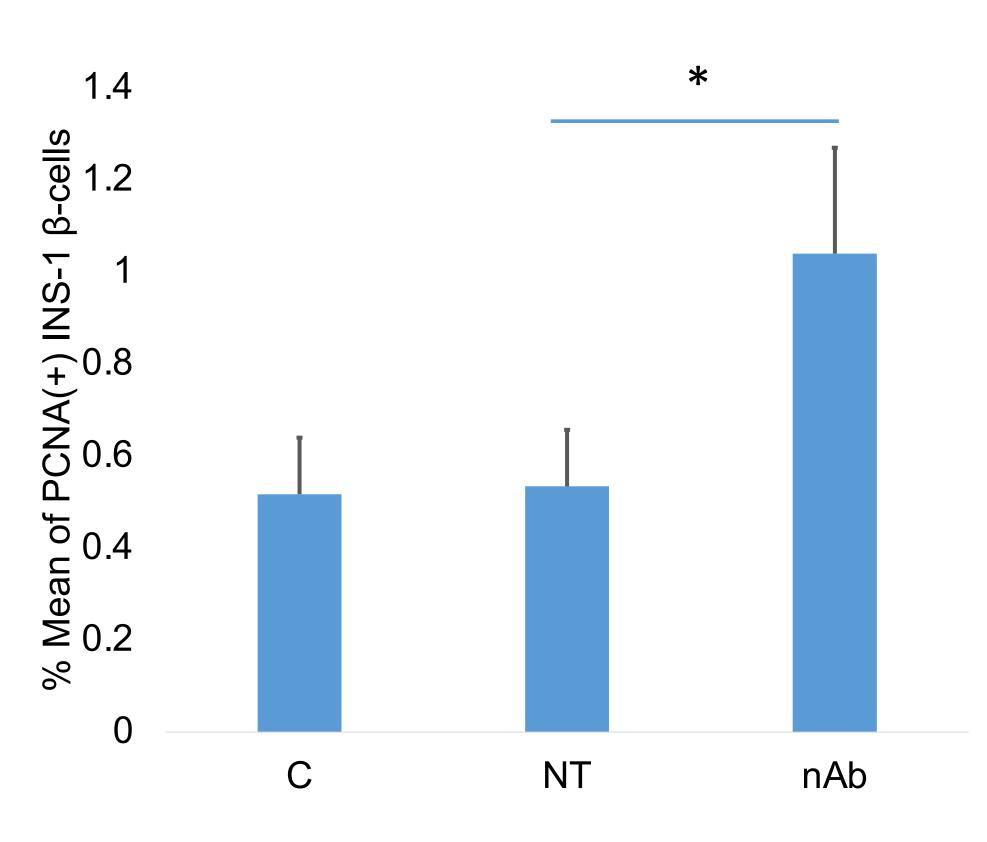




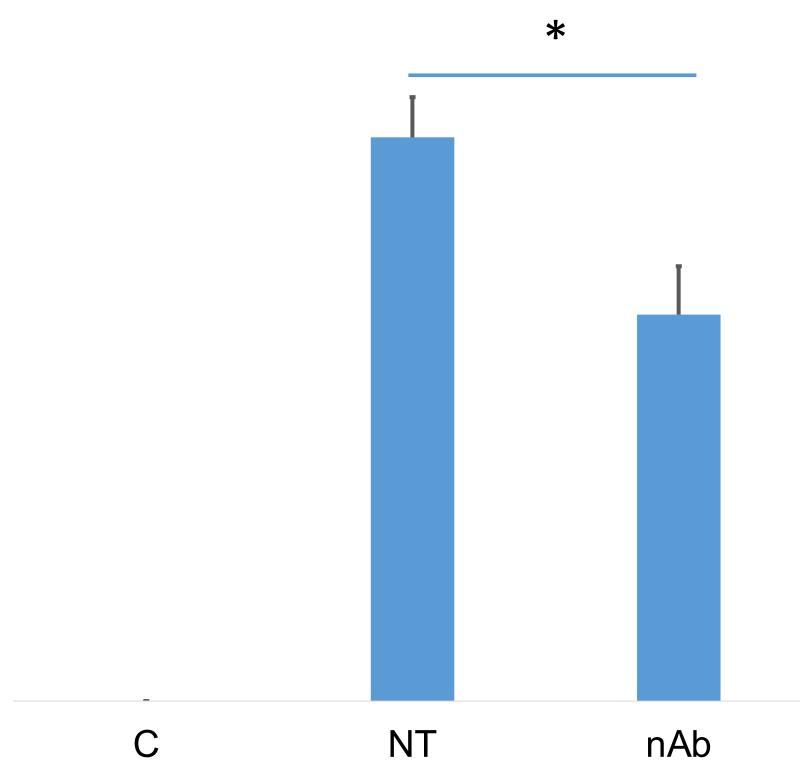
Fig 7. The proportion of amyloid-positive transduced INS-1 ßcells with and without treatment with nAb. Day 0 (C) and Day 7 (nontreated, NT) are shown for comparison. Data are expressed as mean±SEM of three independent studies, \*=p<0.05.

#### CONCLUSION

### ACKNOWLEDGEMENTS



**RESULTS**, continued



Treatment of INS-1 ß-cells with nAb significantly reduced TUNEL-positive and amyloid-positive ß-cells. PCNA-positive ß-cells were also increased post treatment.

• Treatment with nAb significantly reduced intra-cellular amyloid formation, decreased amyloid-induced ß-cell death, and enhanced ß-cell survival (proliferation).

Reducing amyloid formation by blocking IL-1ß signalling may provide an effective approach to decrease loss of ß-cell mass in patients with T2D.

Reducing amyloid-induced IL-1ß signalling may also prove to be of benefit in increasing the longevity of islet grafts patients with T1D.

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